



Ligase Inhibition Workshop

3rd Annual Ligase Targeting Drug Development

Tuesday April 11th 2023

9:00 – 11:00

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Agenda

- Introduction to Ligases (Sumit) – 5 min
- Mechanisms of Inhibition (Sumit) – 5 min
- Ligand-ability of the ligase class (Xevi) – 5 min
- Review of the existing chemical matter (Xevi) – MDM2, VHL, IAP, SOCS2, KEAP1 – 10 min
- HDM2 Structure Based Drug Design (Xevi) – 25 min
- Break – 10 min
- DEL platform overview and advantage (Sumit) – 15 min
- Pellino1 case study (Sumit) – 25 min
- Q&A

Ubiquitin Ligase are Mediators of Cellular Signaling and Homeostasis



Nurix Drugs Engage Ligases for the Treatment of Cancer

Targeted Protein Modulation: $TPM = TPD + TPE$

A Powerful
Cellular System



Targeted Protein
Elevation
(TPE)

Harness ligases
to decrease
specific protein levels

Inhibit ligases
to increase
specific protein levels

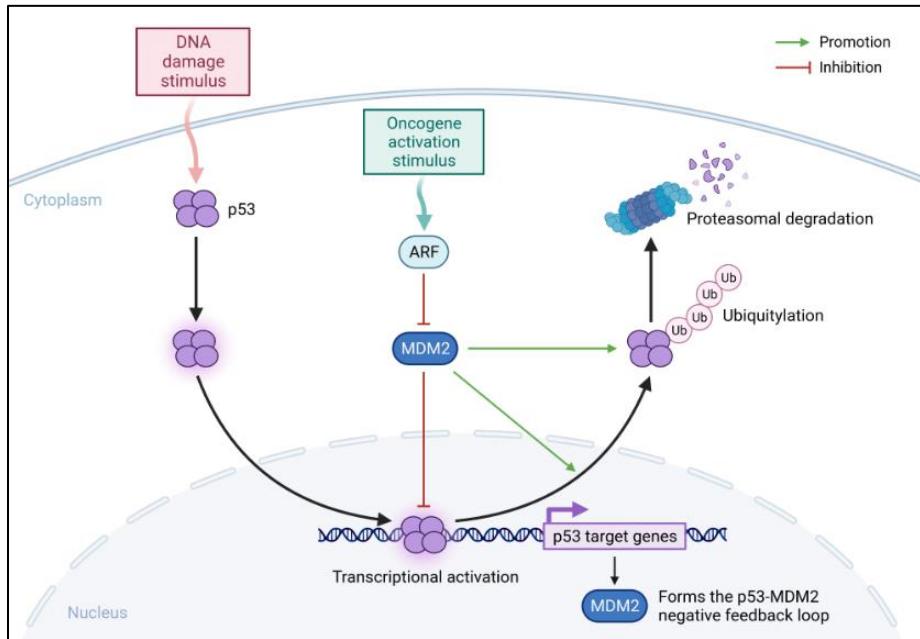
Targeted Protein
Degradation
(TPD)

Ubiquitin is ligated to
target proteins to tag
them for degradation by
the proteasome

Ligase Inhibition as a Therapeutic Strategy

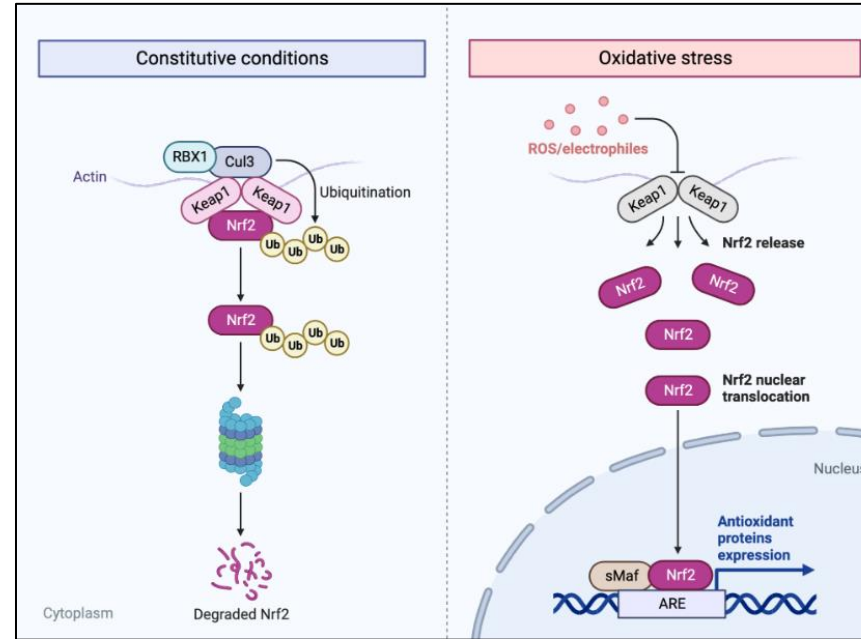
- E3 ligases control cellular protein abundances in response to various stimuli
- They are key regulators of disease relevant cellular pathways

Oncogenic Signaling (e.g., MDM2)



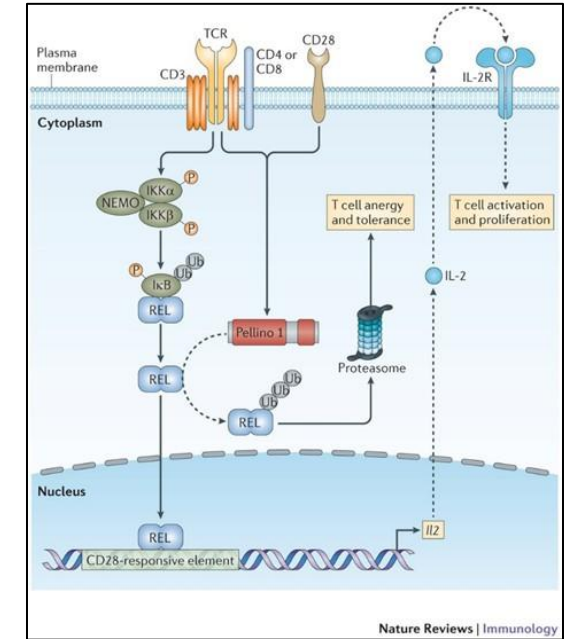
Biorender

Redox Signaling (e.g., KEAP1)



Biorender

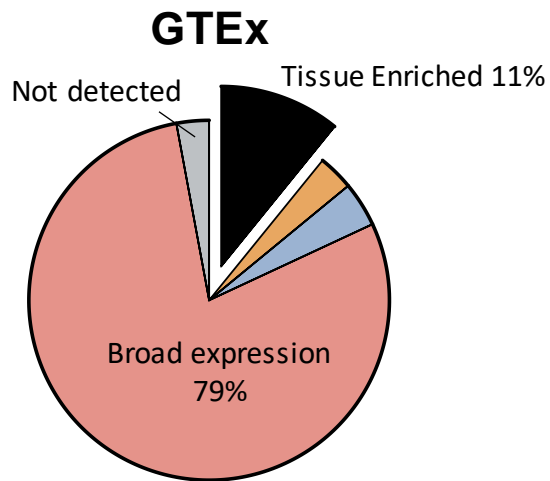
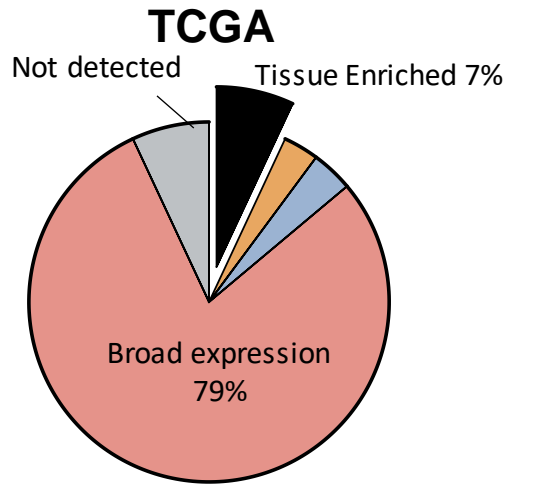
Receptor Signaling (e.g., Pellino1)



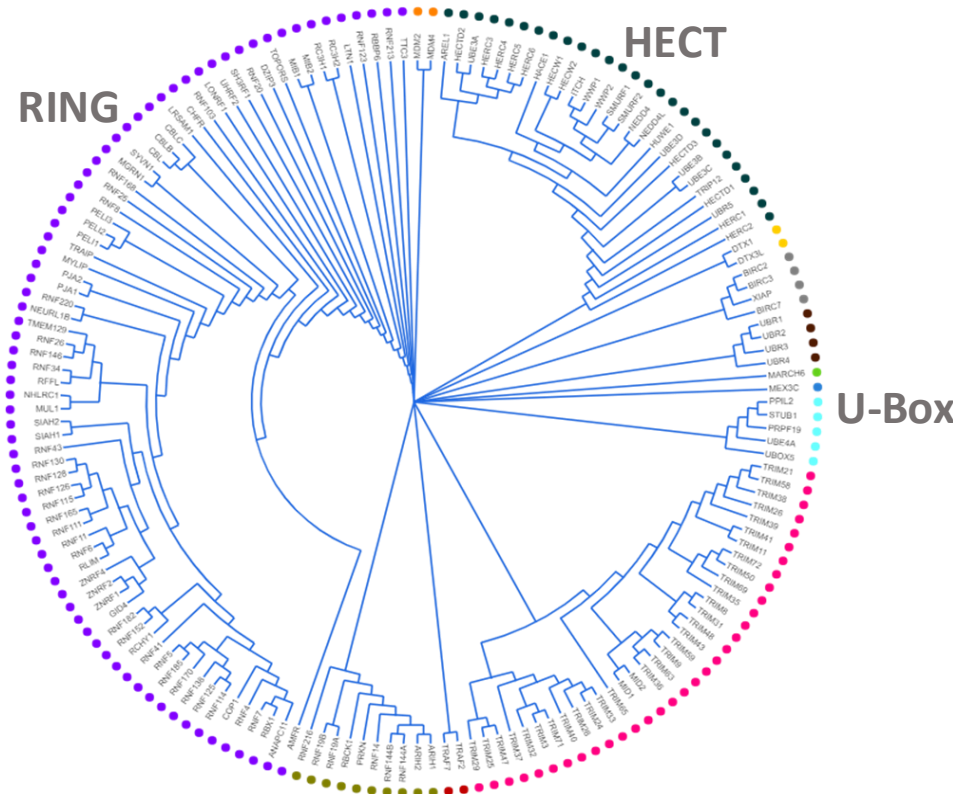
Nature Reviews | Immunology
Moynagh PN, Nat Rev Immuno 2014

The Ubiquitin Ligase Family is Diverse with Over 600 Proteits

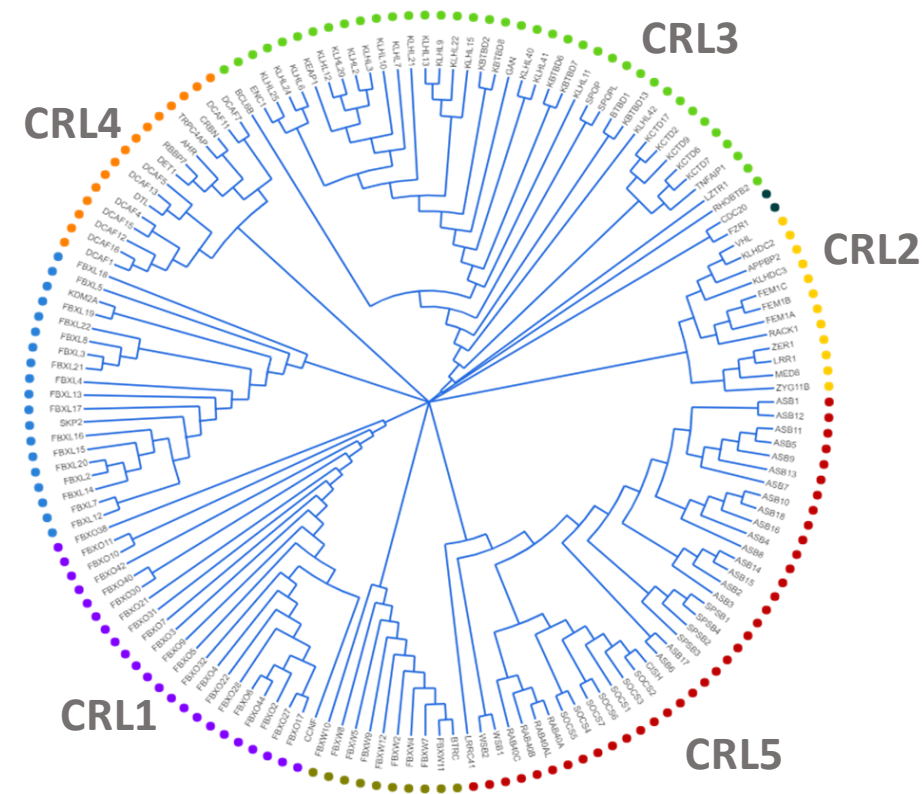
- Diverse family of proteins that are broadly expressed in most tissues
- Classified into single chain or multimeric ligases based on the functional assembled state
- Sub-classified based on their conserved motif or domains



Single Chain E3 ligases

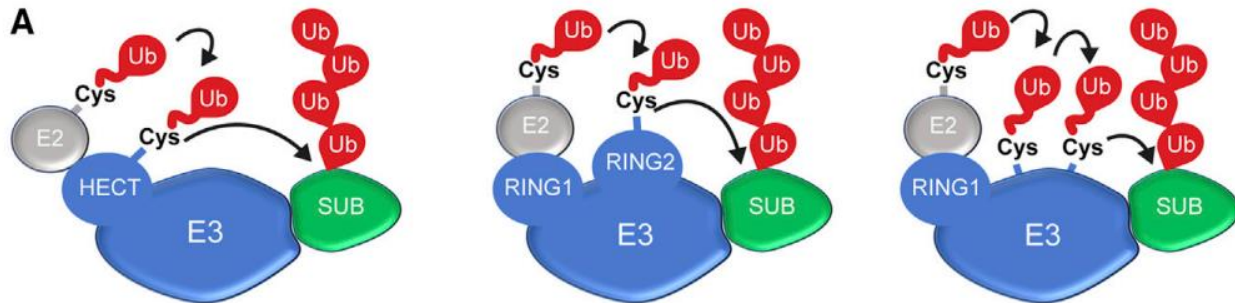


Multimeric E3 ligases Substrate Binding Modules



SGC UbiHUB

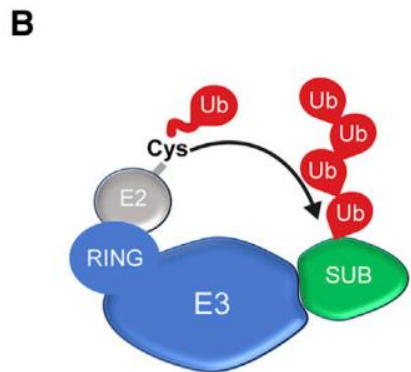
E3 Ligases assemble diverse ubiquitin chains on their substrates



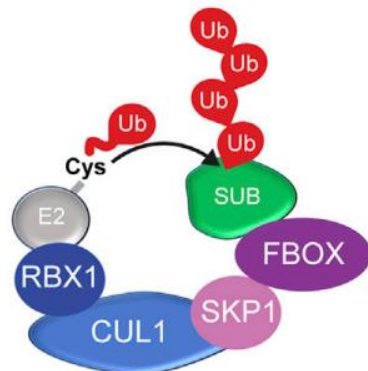
HECT E3 ligase
i.e. *UBR5, NEDD4*

RBR E3 ligase
i.e. *Parkin, ARIH1*

RCR E3 ligase
i.e. *MYCBP2*



RING E3 ligase
i.e. *MDM2, cIAP1, UBR1*



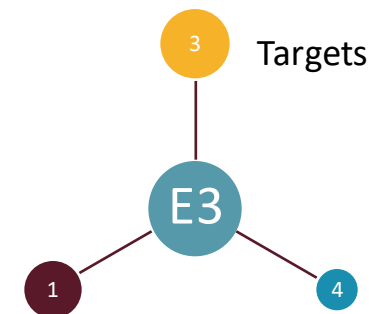
CULLIN RING E3 ligase
i.e. *SCF, CUL2^{VHL}, CUL4^{CRBN}*

- Most E3 ligases lack a well-defined active site
- They work by inducing proximity between the target protein and activated E2~ubiquitin complex
- Ubiquitin forms a covalent bond with a lysine side chain on the target protein. Poly-ubiquitin chain is assembled when additional ubiquitin are attached to multiple the previous ubiquitin leading to a diversity of chain topologies
- Polyubiquitin chain binding receptors lead to distinct outcomes of the modified protein (e.g., Lys-48 linked linear or branched chains are effectively recognized by the proteasomal degradation complex)

Broad acting ligases

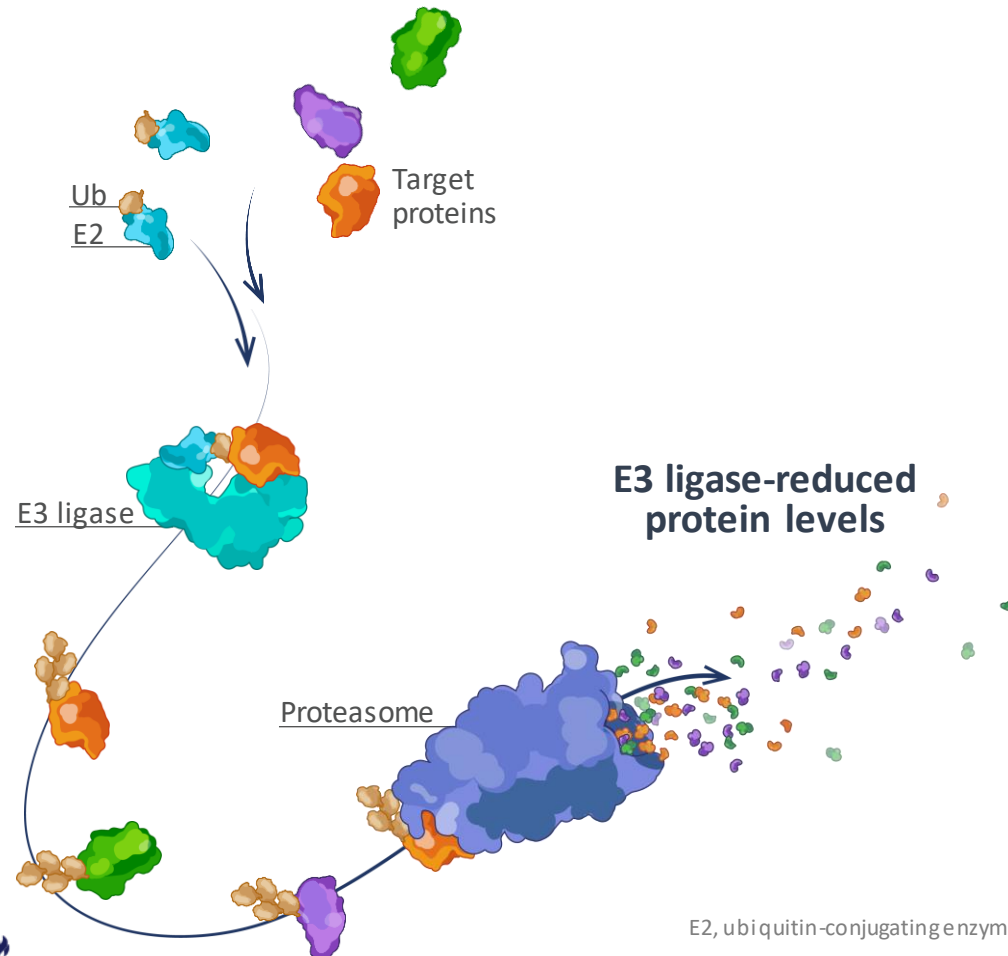


Narrow spectrum ligases

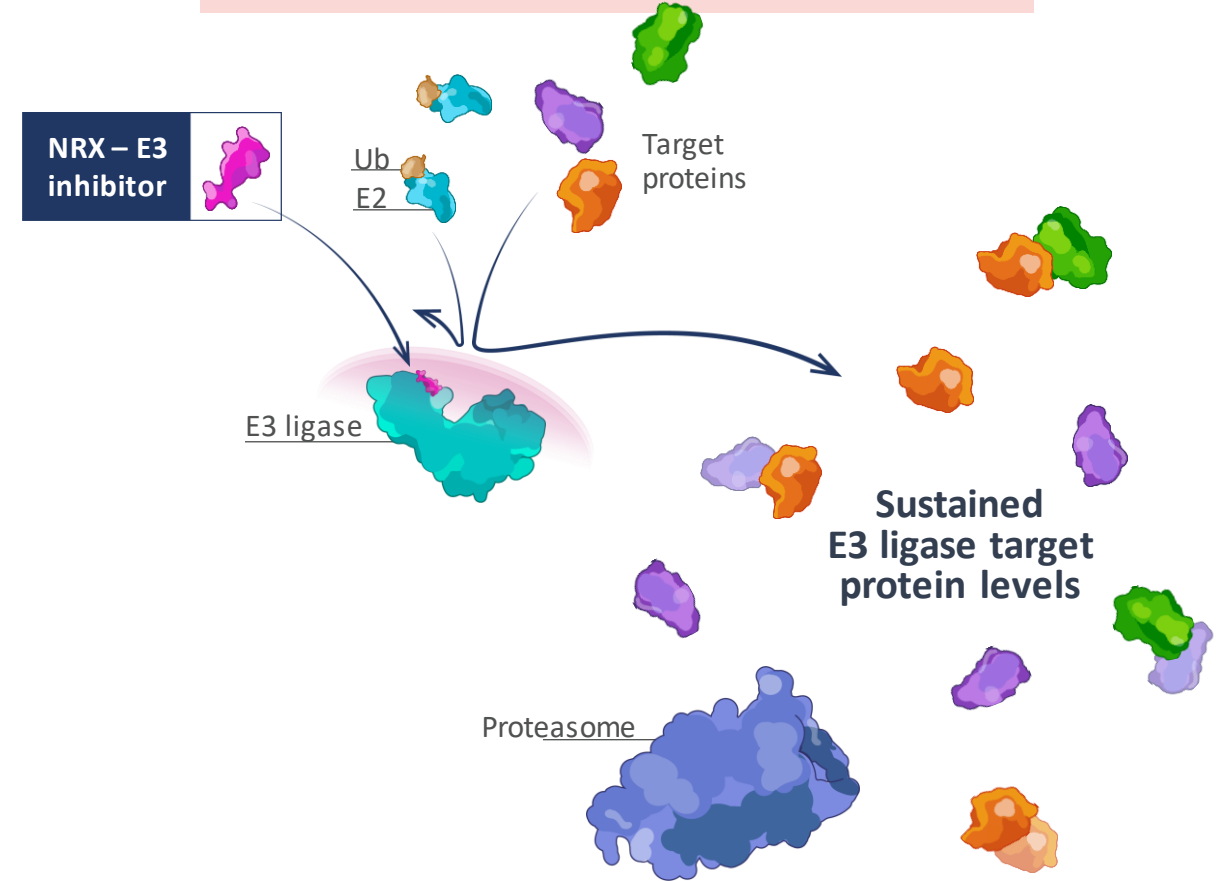


One E3 Ligase Controls Multiple Target Proteins and Pathways

**Native turnover –
Low-protein abundance**



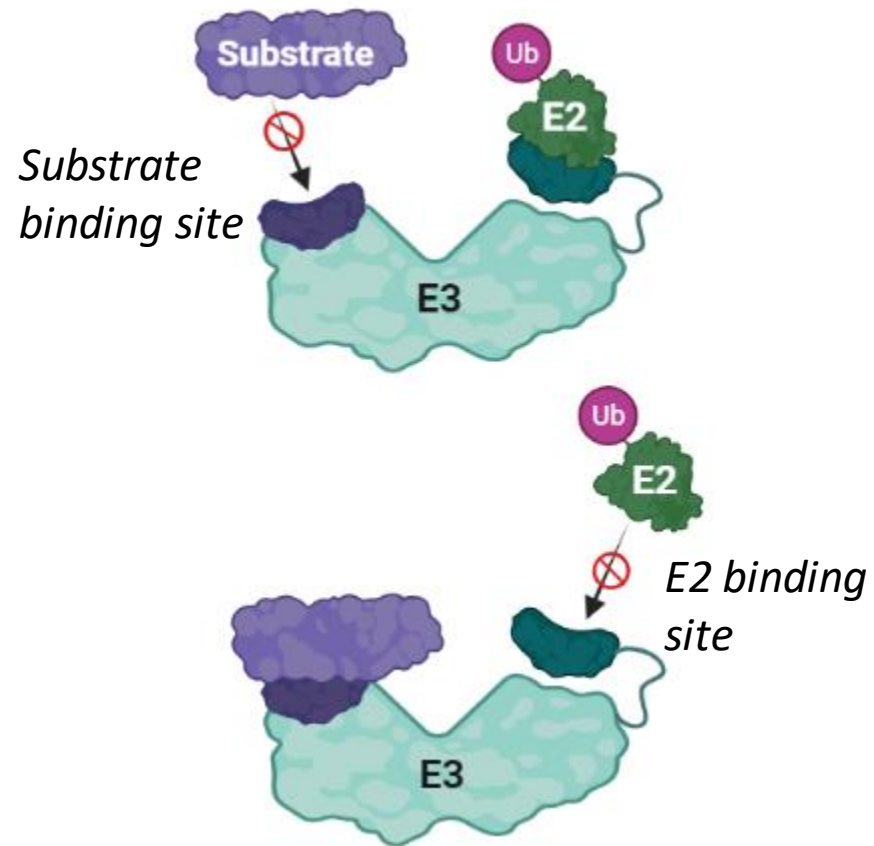
**E3 Inhibitor-induced
High-protein abundance**



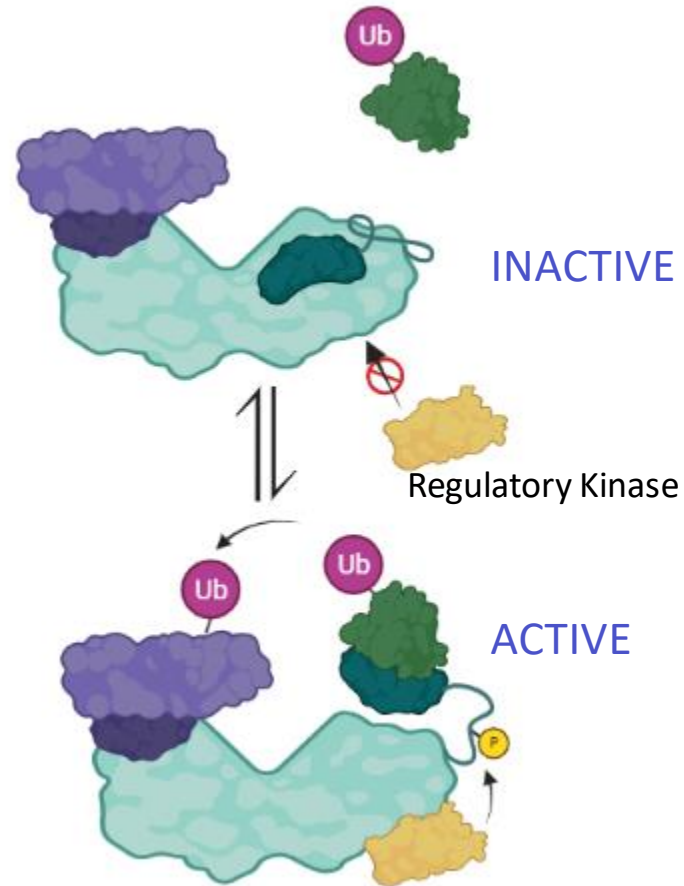
E2, ubiquitin-conjugating enzyme; E3, ubiquitin ligase; NRX, Nurix; Ub, ubiquitin

Mechanisms Targeted by Small Molecule Inhibitors of E3 Ligase

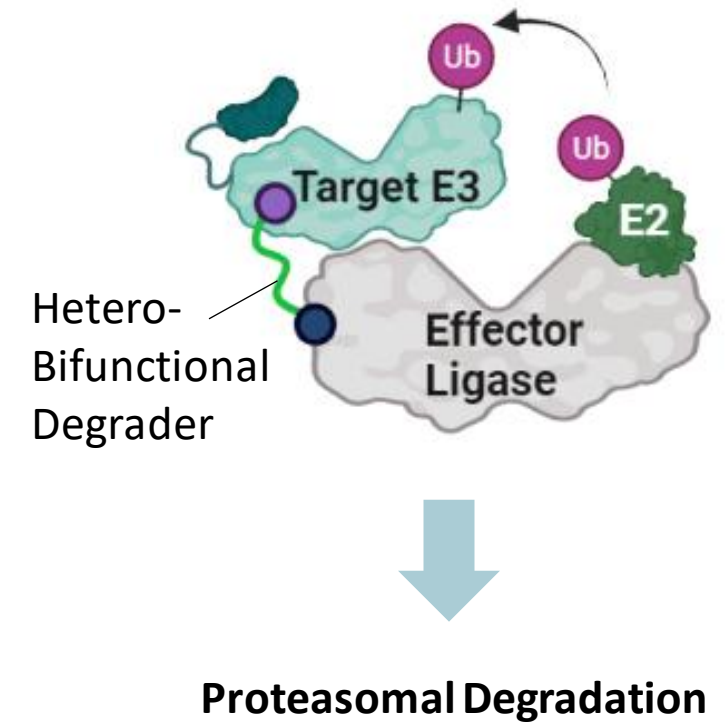
Protein-Protein Interaction



E3 Ligase Activation



E3 Ligase Degradation

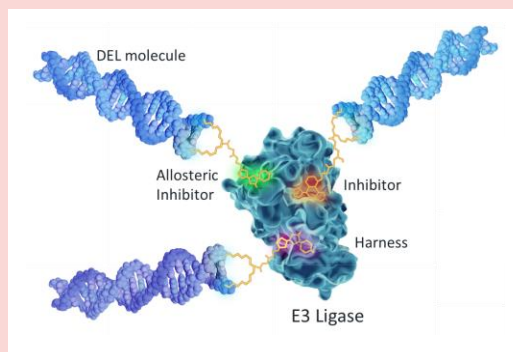


Enabling E3 Ligases Ligand Discovery with DNA-Encoded Library Screening



Nurix's Integrated Protein Modulation Discovery Platform

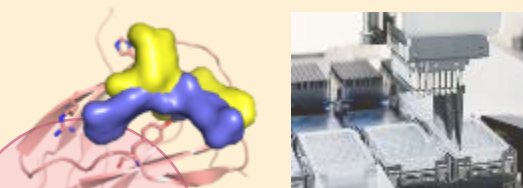
DEL Discovery



> 5 billion drug-like compounds that can be easily screened against hundreds of proteins to identify starting points for therapeutic discovery

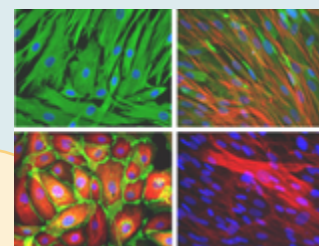
Supported by strength in Protein Sciences

Rational and Empirical Chemistry



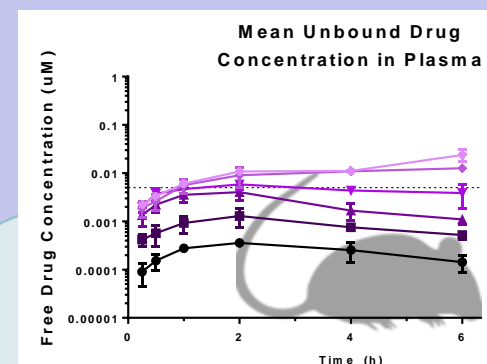
Structure Based Drug Design combined with chemistry automation enables broad exploration of lead-like chemical space for each program

Direct-to-Cell Biology Capabilities



High throughput cellular assays monitor protein levels and biological phenotypes to assess impact on biology

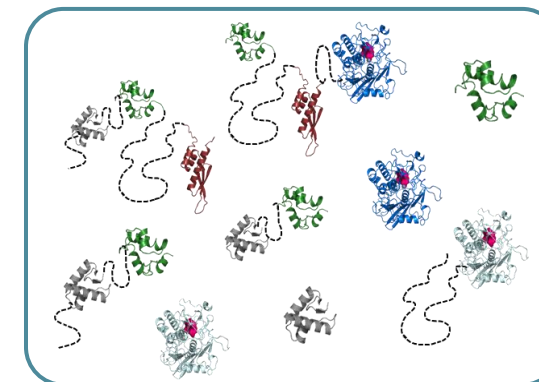
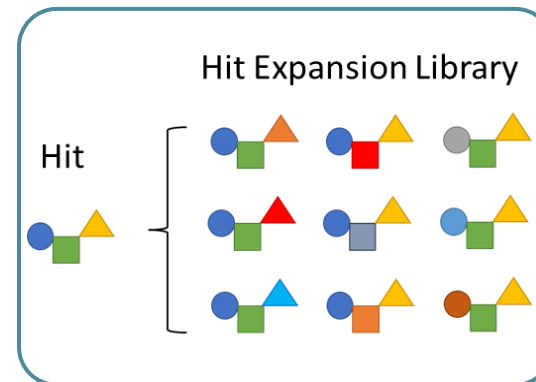
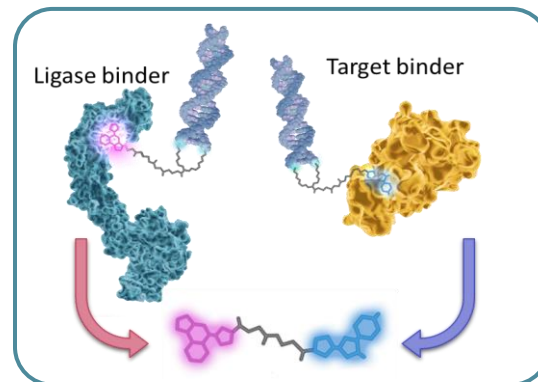
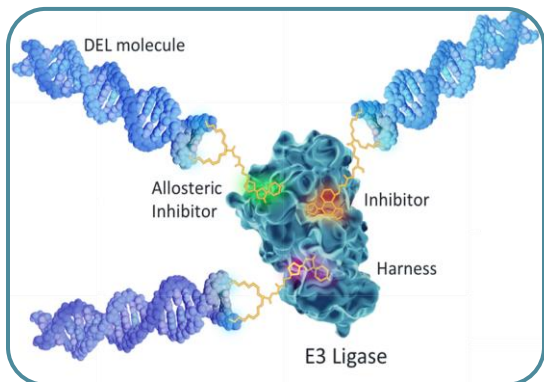
Scaled Screening for *in vivo* exposure



Capacity to screen for ideal *in vivo* drug exposure profile and assess impact on disease biology

Expertise in Oncology and immuno oncology

Why DNA Encoded Libraries?



Affinity-based screening is MoA agnostic – for E3 ligases we can identify ligands for TPD and inhibitors for TPE from the same screen

DNA attachment provides initial handle for bifunctional molecule synthesis

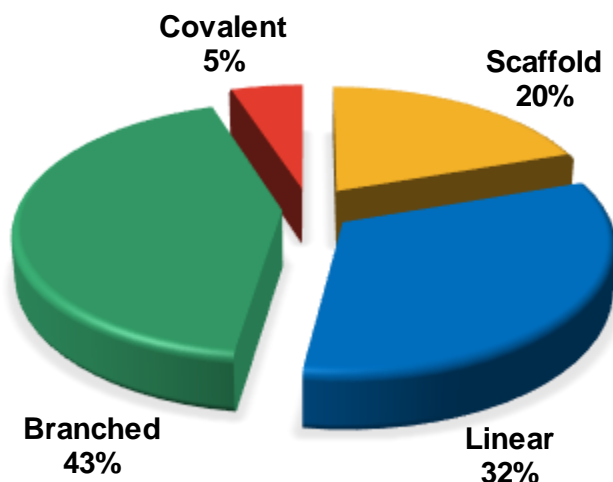
Combinatorial design enables rapid hit follow up and optimization

Low capital investment and per screen cost allows for a broad exploration of target and chemical space

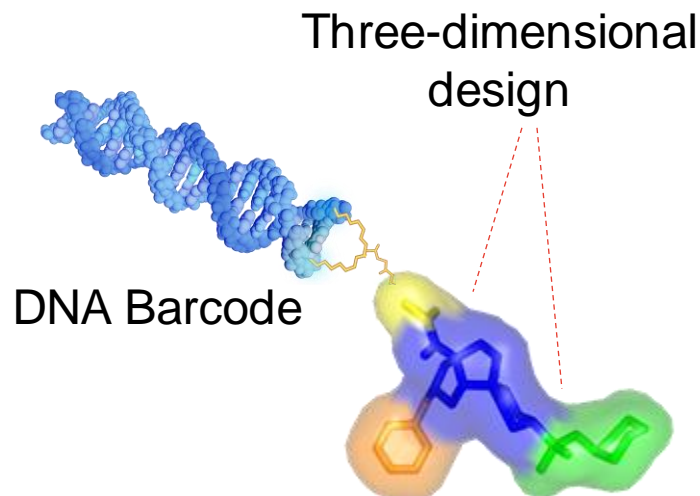
Custom Scaffold-Based DELs Enable Nurix To Identify Binders to Challenging Protein Surfaces

Nurix DEL Collection

- >5 billion unique structures
- Includes proprietary, 3D complex, custom scaffolds

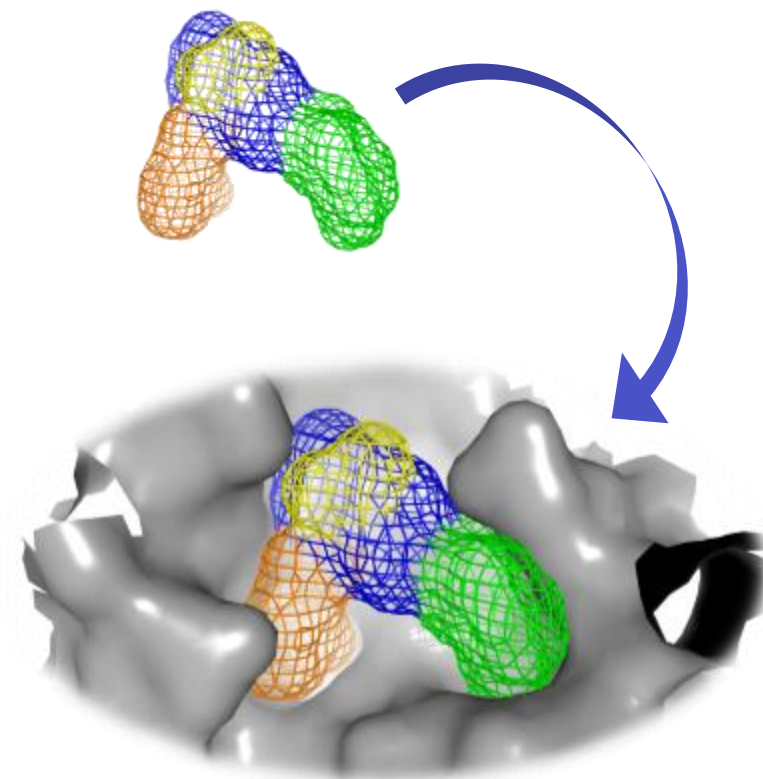


Scaffold Libraries Proving Essential for Delivering Ligands for “Undruggable” Targets (sole source of hits for 75% of these targets)



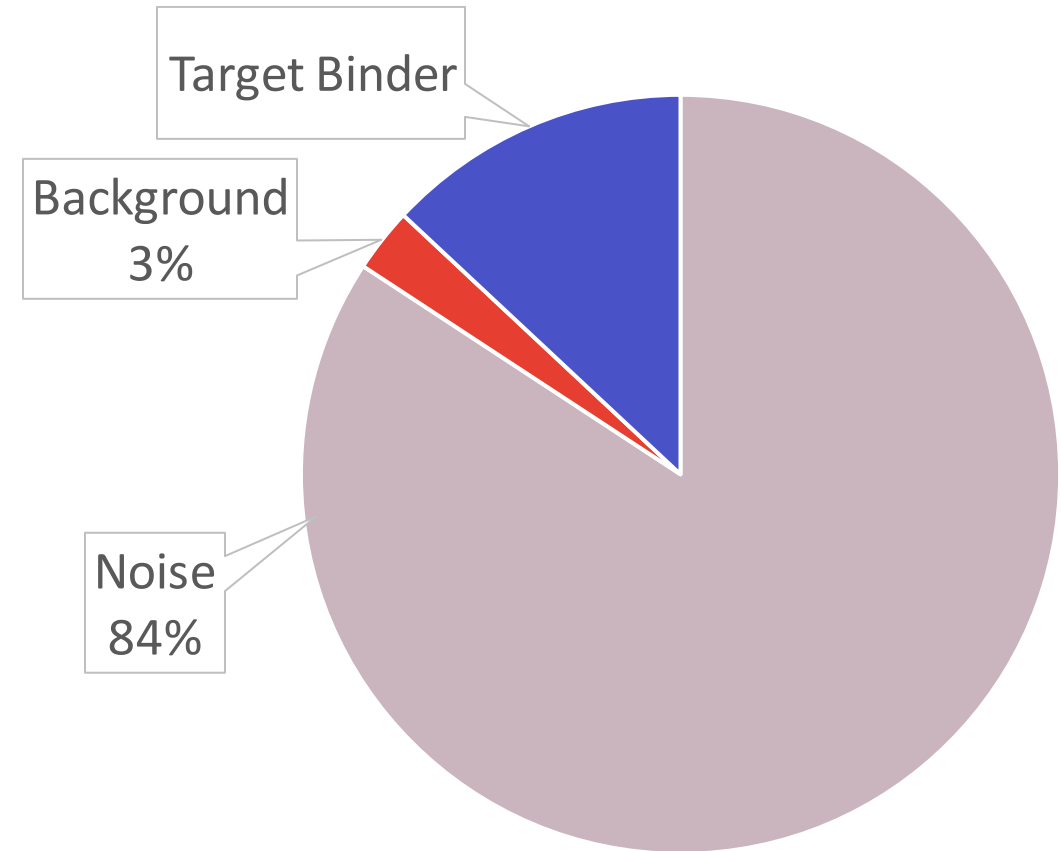
Our proprietary scaffold DELs provide unique geometry and high sp³ character, allowing molecules to achieve optimal pocket fit

Nurix scaffold designs show high pocket complementarity



Composition of DEL Screening Outputs

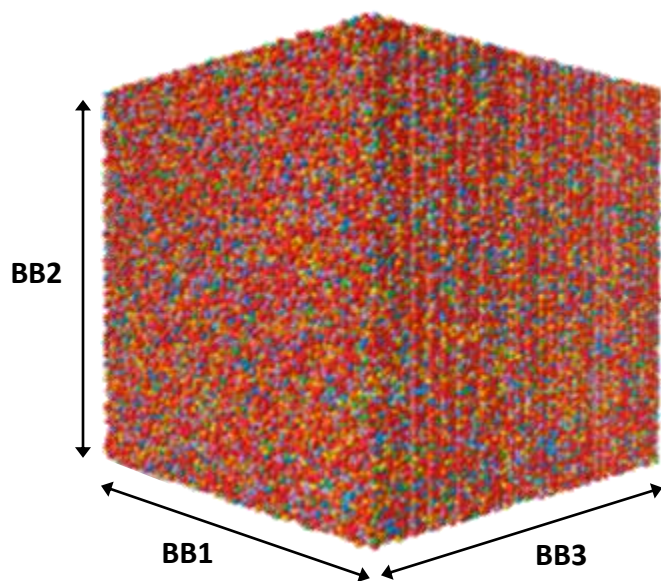
- Most of the DNA-linked compounds sequenced at the end of a selection are noise or background (matrix binders, non-specific protein binding, other enrichment not specific to the target)
 - Noise can be eliminated by experimental (replicates) OR analytical (thresholding) methods
 - Elimination of background signal requires the combination of experimental AND analytical methods.



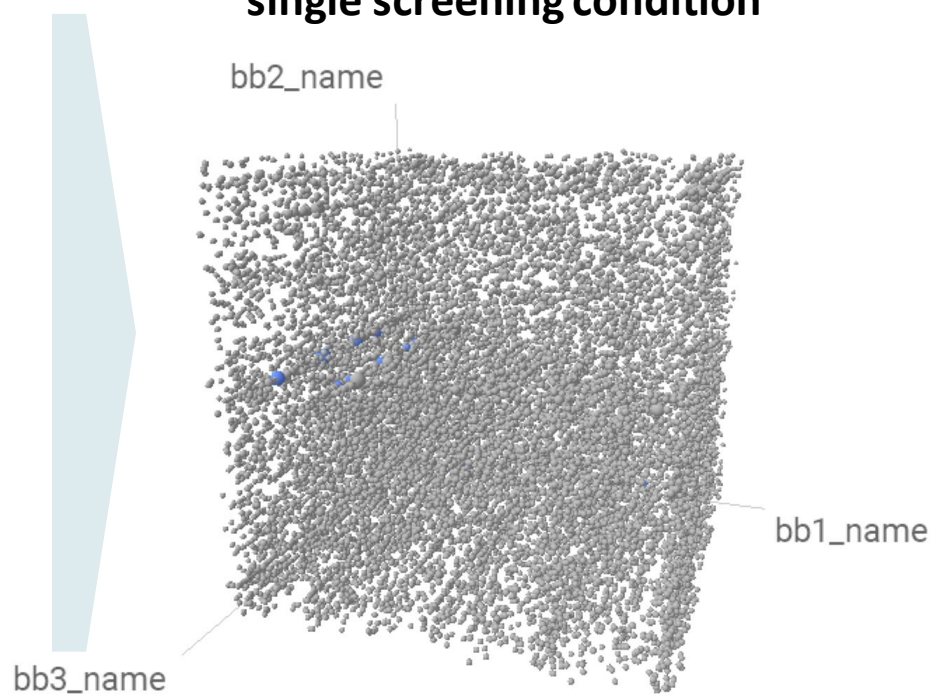
De-noising Example – VHL Replicates

- Noise by its nature is not reproducible, but real binding events are.

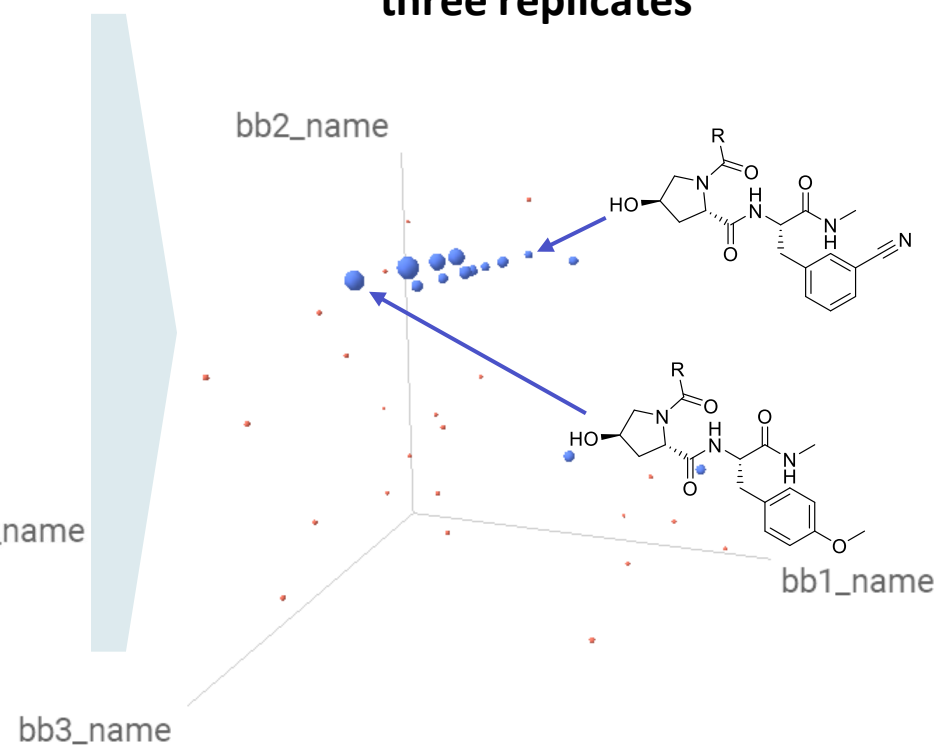
Nurix DEL Collection
>5 billion unique structures



All ligands present in a single screening condition

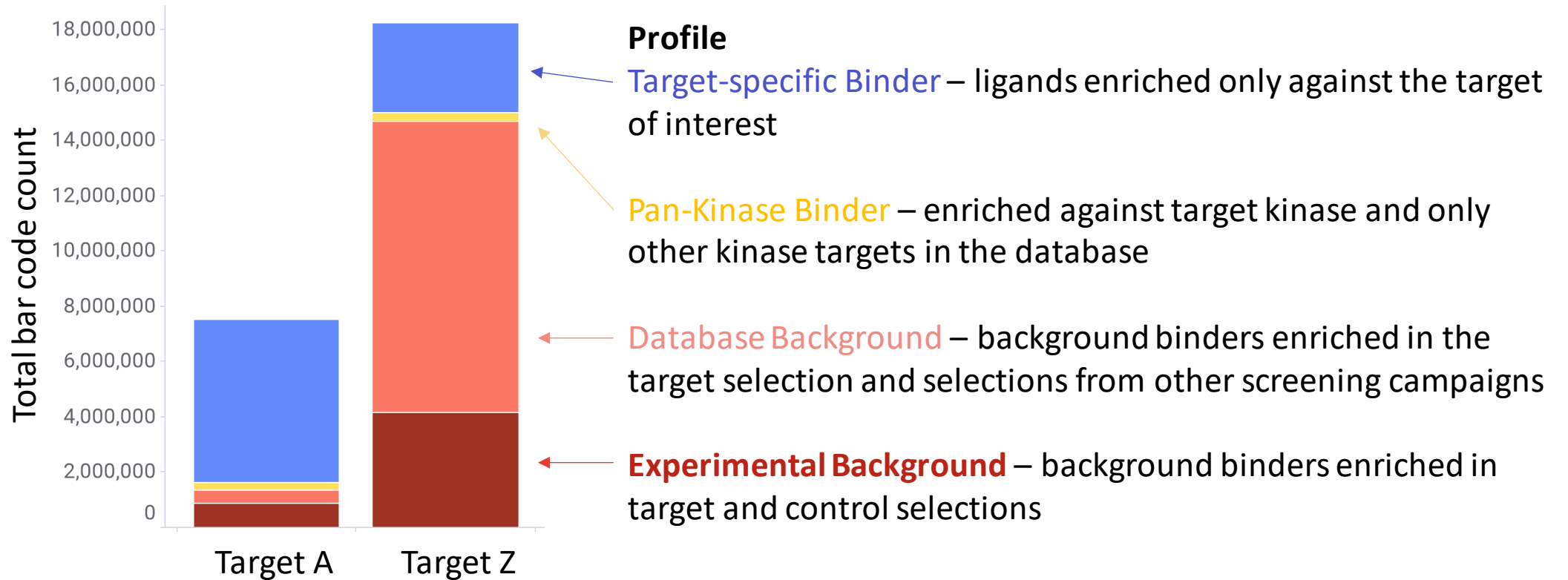


All ligands present in all three replicates



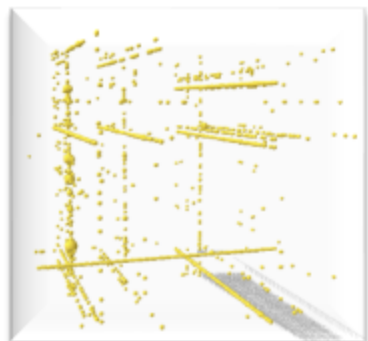
A Robust Database Is Necessary for Effectively Identifying Background

- A combination of experimental AND analytical methods are required to effectively eliminate background.
- Not all background binders are identified in control screens.
- The capacity of the platform enables screening across many targets, which powers a database that can effectively remove background binders and identify selective (and non-selective) target binders.

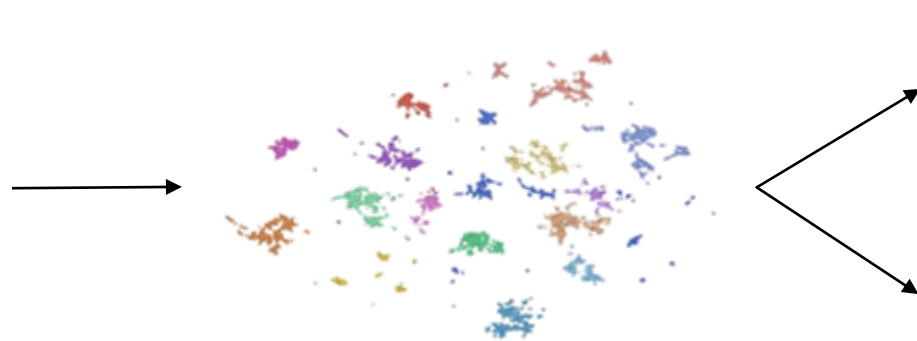


Wading Through the Data - Nurix's Analysis and Follow Up Pipeline Is Designed To Access Broad Chemical Space

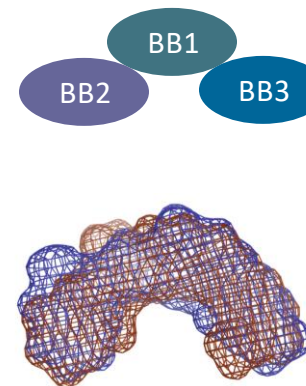
Large complex data sets require automated solutions to accelerate hit ID



DEL Screen and filtering for target-specific binders



Automated Structure Analysis and Clustering



Hit Resynthesis (on- and off-DNA)

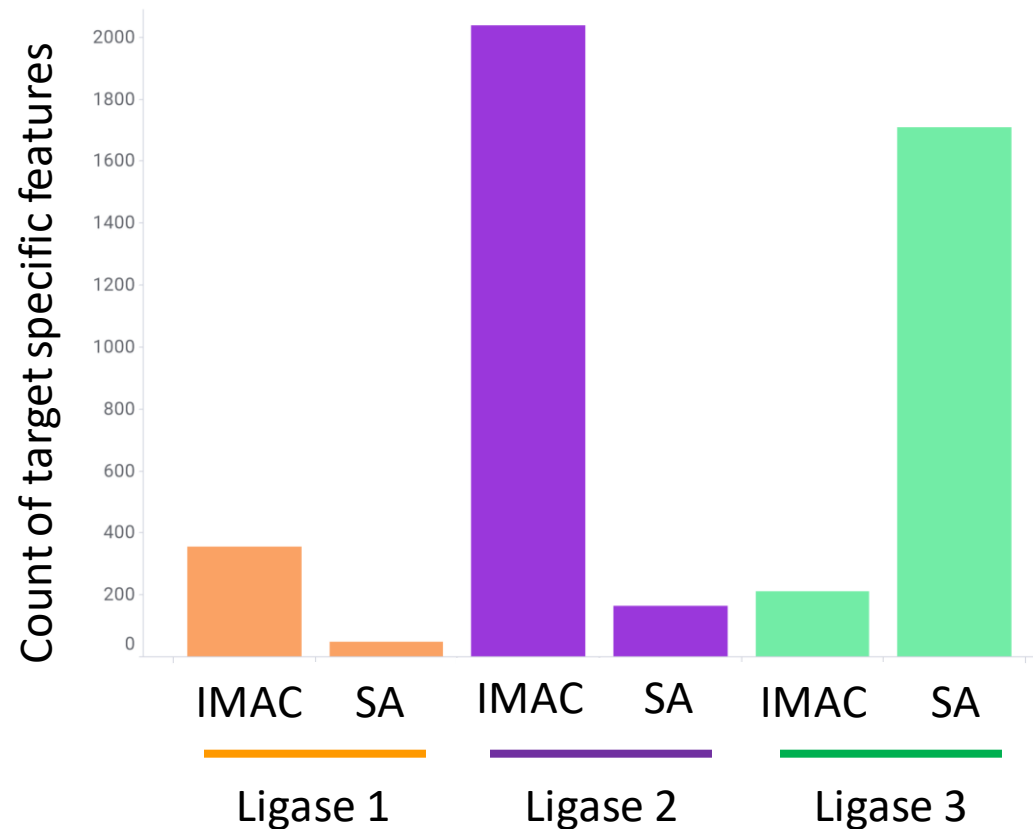
Machine Learning and Similarity Virtual Screening

Follow up	Source	Volume	Hit Confirmation Assay
Off-DNA	Single compound synthesis	10s	SPR (Quantitative)
On-DNA	Parallel Synthesis of single recipes	100s	ASMS (Qualitative)
ML/Similarity	Catalog order	100s	ASMS then SPR (Quantitative)

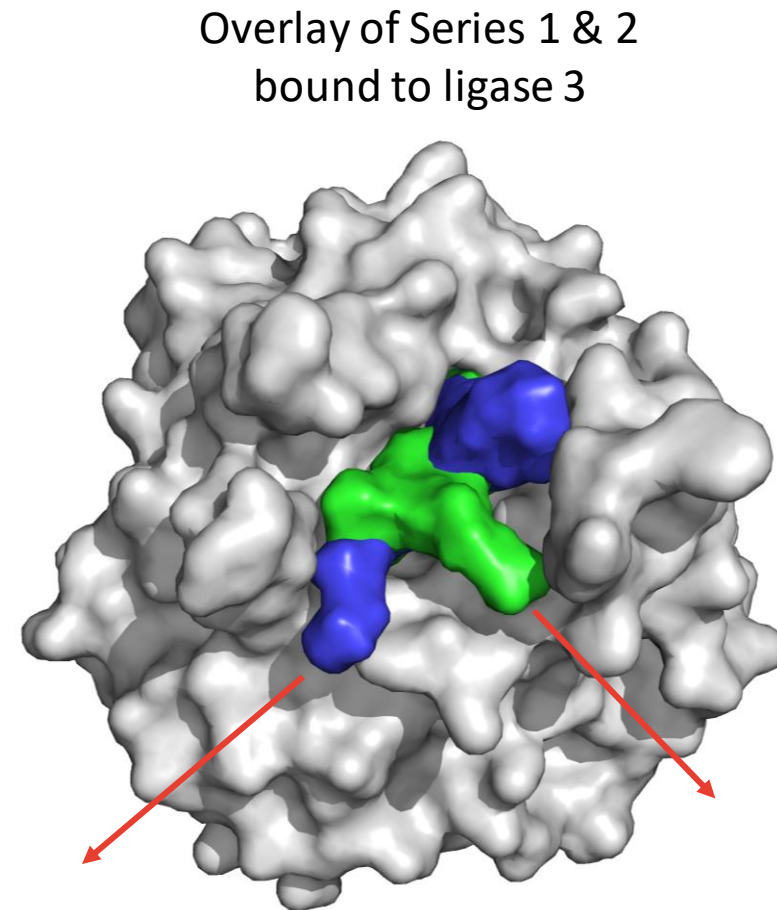
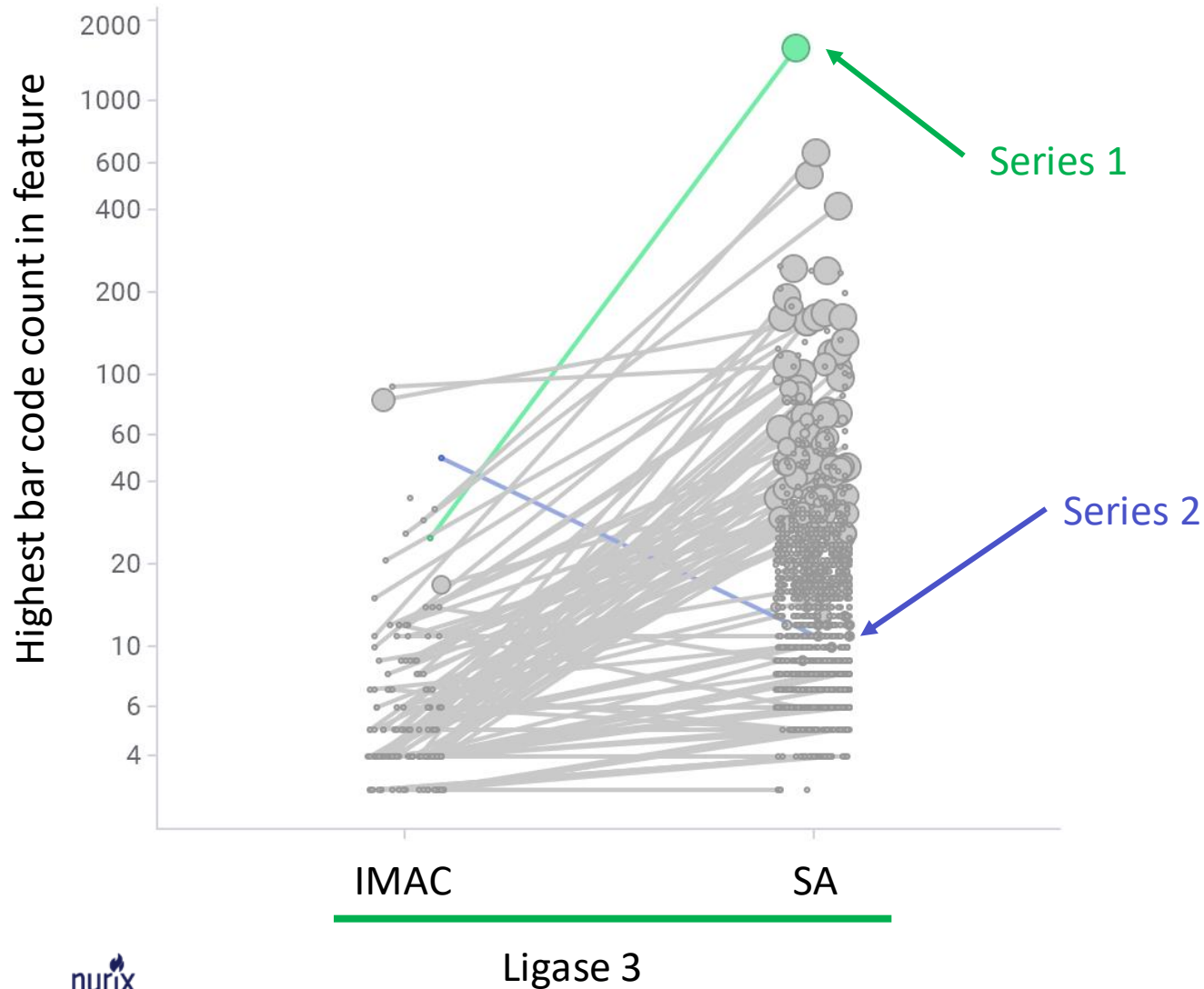
Screening and Follow Up Capacity – Finding the Most Productive Spaces for Novel Targets

- Screening multiple ligases in parallel, with multiple constructs and tags for each ligase
- Nurix routinely screens multiple target constructs immobilized through different matrices
 - The most productive construct/matrix combinations needs to be determined empirically

Example – three ligases screened in parallel using Immobilized metal affinity (IMAC) and streptavidin (SA) beads



Broad Follow Up Maximizes the Opportunities from DEL Screens



Unprecedented ligase with two confirmed vectors for bifunctional attachment

Conclusions

- DEL provides significant advantages as a ligand discovery platform for targeted protein modulation
- These advantages can only be realized when coupled to high-quality, well-validated target proteins and a diverse collection of libraries
- Leveraging the low cost per screening condition and the ability to broadly scan the chemical space of hits are key to maximizing the productivity of the platform
- Assembling a comprehensive database of screening results from a broad exploration of target space is key to navigating through the data to find the highest quality hits

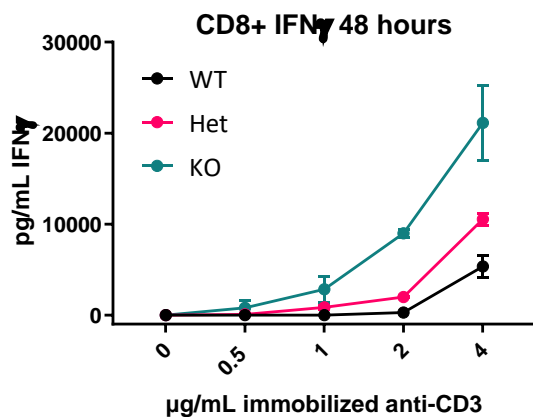
Pellino1 Case Study



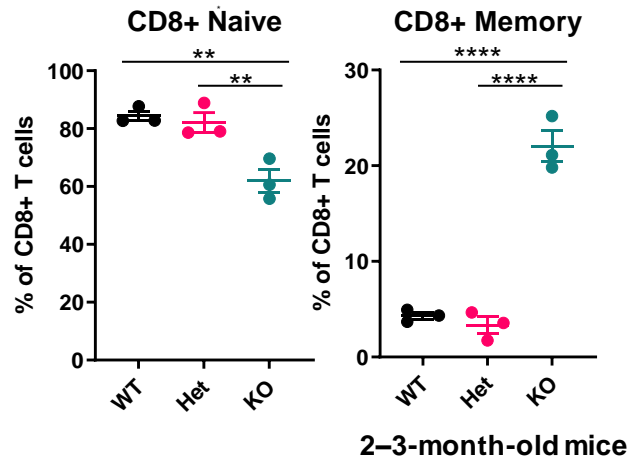
Pellino1 is an Immuno-oncology Target

- Pellino1 is an E3 ligase which is a negative regulator of T cell activation
- Therapeutic hypothesis: Degradation of Pellino1 will result in an anti-tumor response by increasing T cell activation
- Peli1 knockout mice display phenotypes consistent with therapeutic hypothesis:
 - T cells display hyperactivation when profiled ex vivo
 - T cells display increased memory markers in vivo
 - Knockout mice display a tumor growth inhibition phenotype

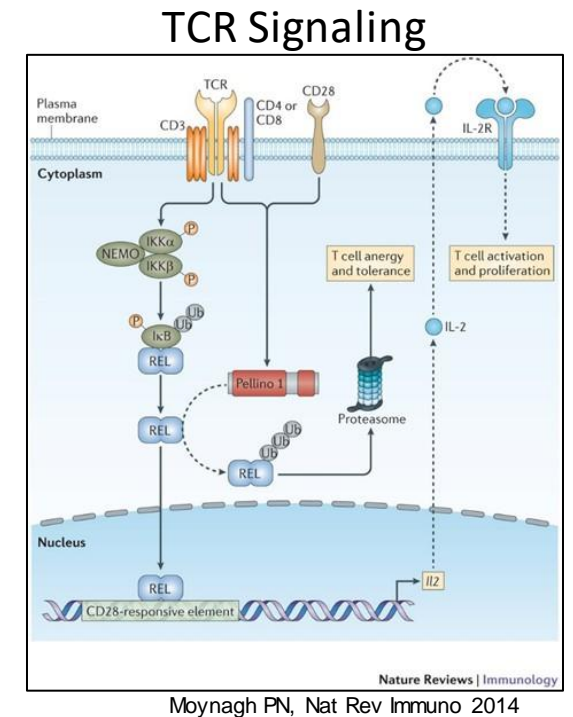
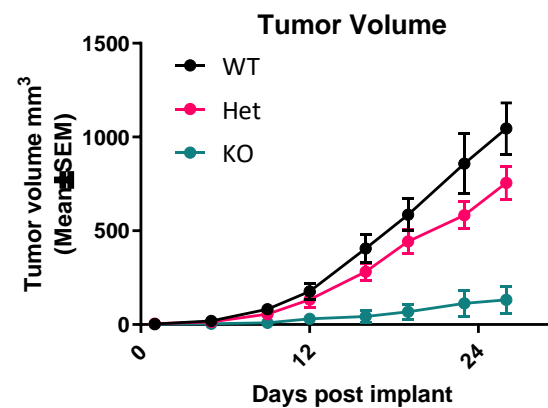
Enhanced Cytokine Secretion



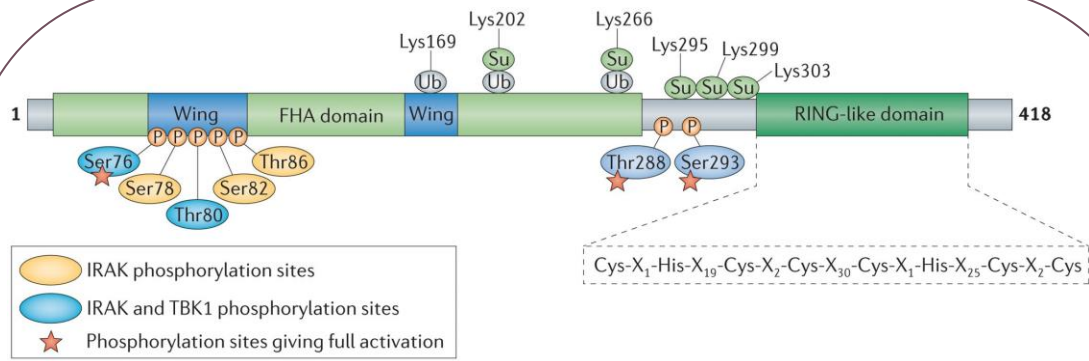
Increased Memory T Cells



Tumor Growth Inhibition

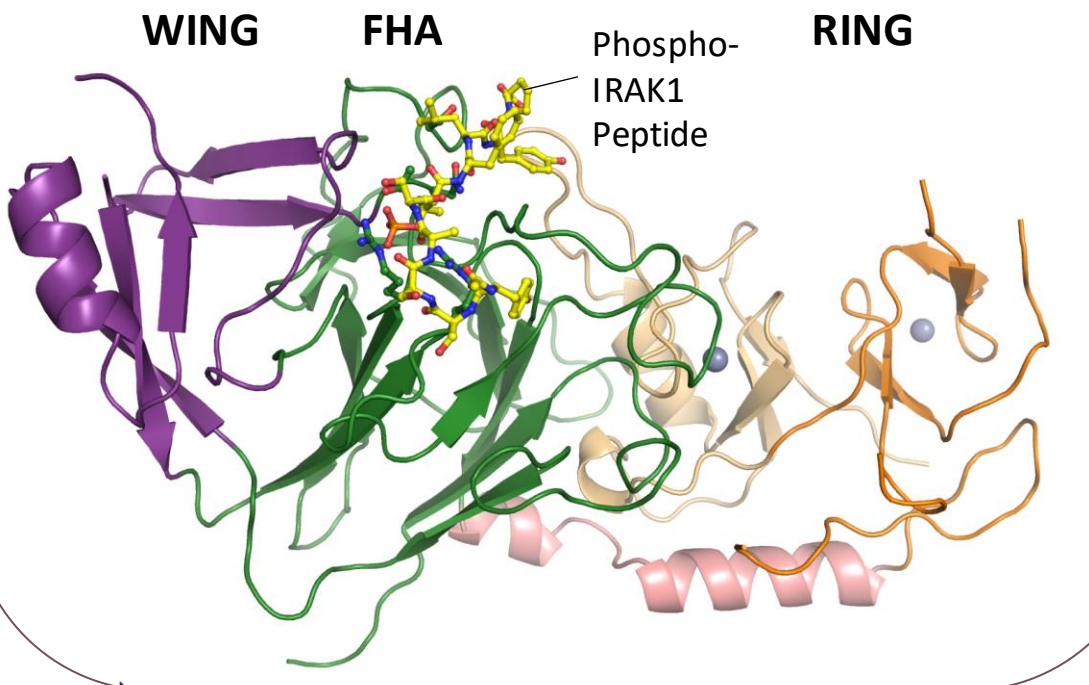


Pellino1 is a RING E3 ligase activated by phosphorylation

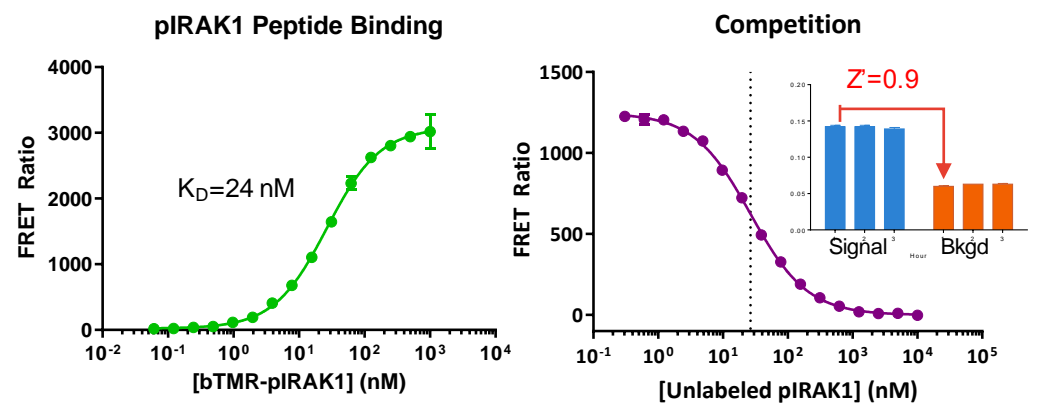


● IRAK phosphorylation sites
● IRAK and TBK1 phosphorylation sites
★ Phosphorylation sites giving full activation

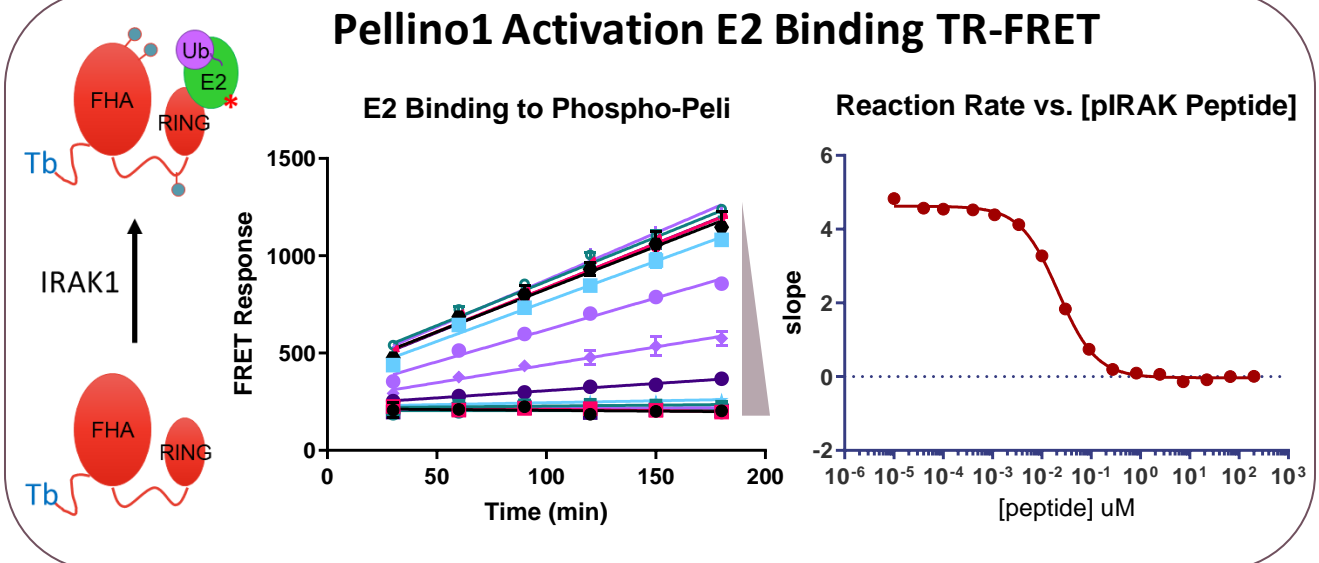
Nature Reviews | Immunology



IRAK1 Phospho-Peptide Displacement TR-FRET

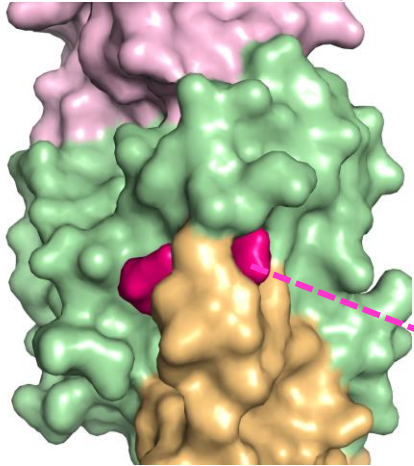


Pellino1 Activation E2 Binding TR-FRET

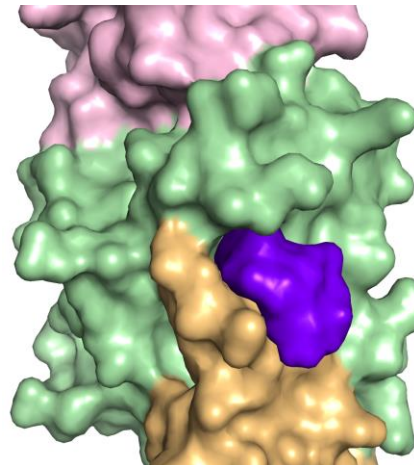
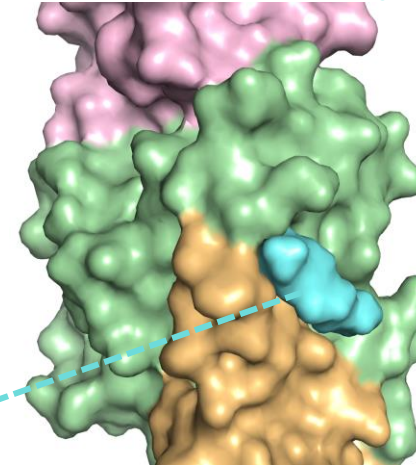


Multiple Hit Finding Approaches Yield Pellino1 Binders

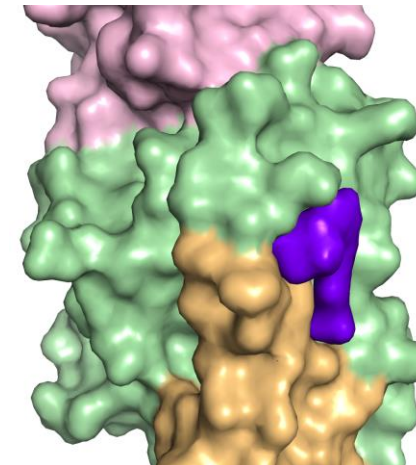
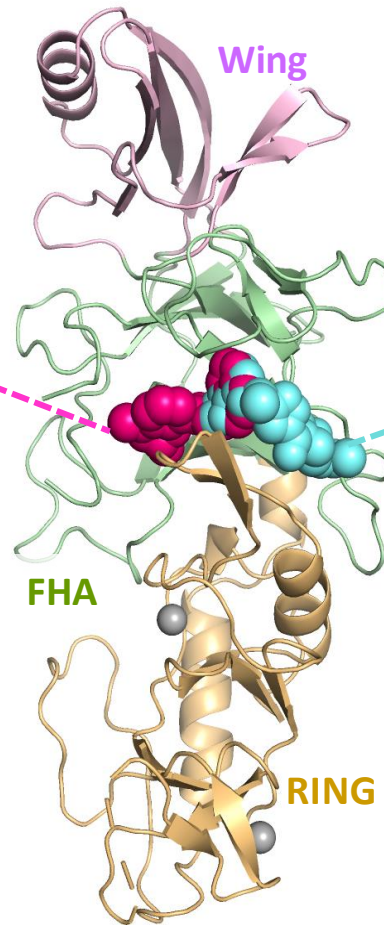
Derived from HTS hit (series 1)



Derived from HTS hit (series 2)



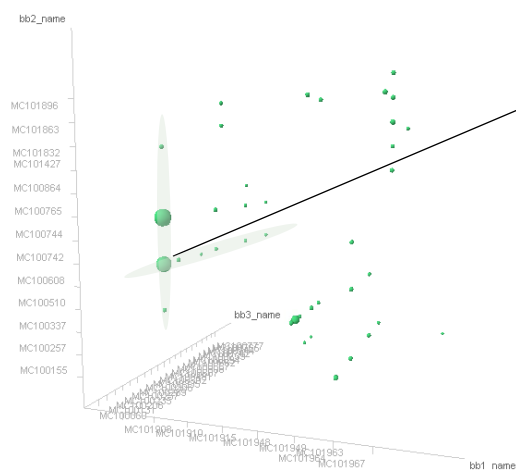
DEL hit (Series 3)



DEL hit (Series 4)

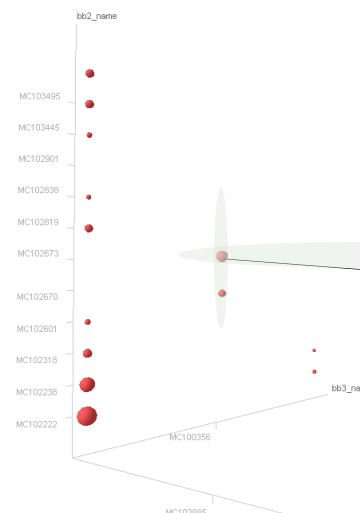
Two Series Confirmed as Pellino1 Binders from DEL Screen

DEL Output



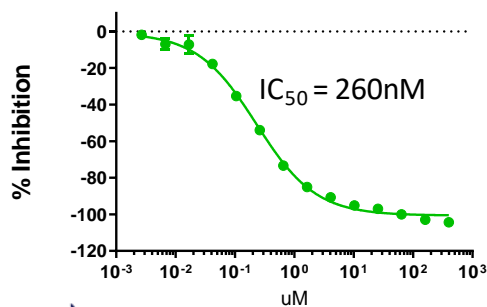
NRX-1
 MW = 542.6
 cLogP = 2.1
 LE = 0.24
 LipE = 4.8

DEL Output

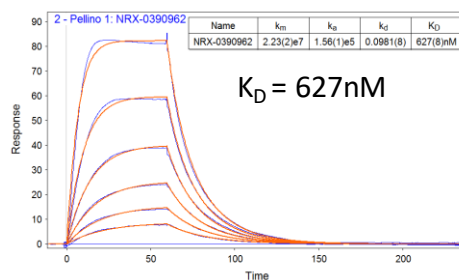


NRX-2
 MW = 472.6
 cLogP = 3.6
 LE = 0.25
 LipE = 2.7

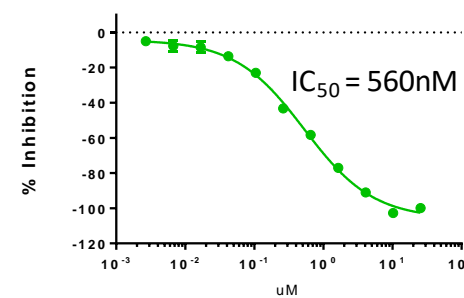
Peptide Binding Inhibition (FRET EC₃₀)



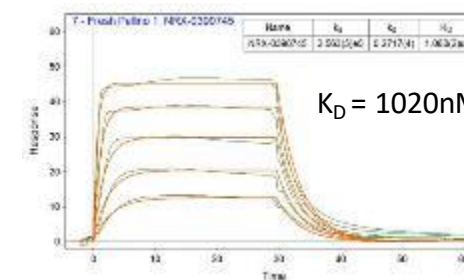
Surface Plasmon Resonance



Peptide Binding Inhibition(FRET EC₃₀)

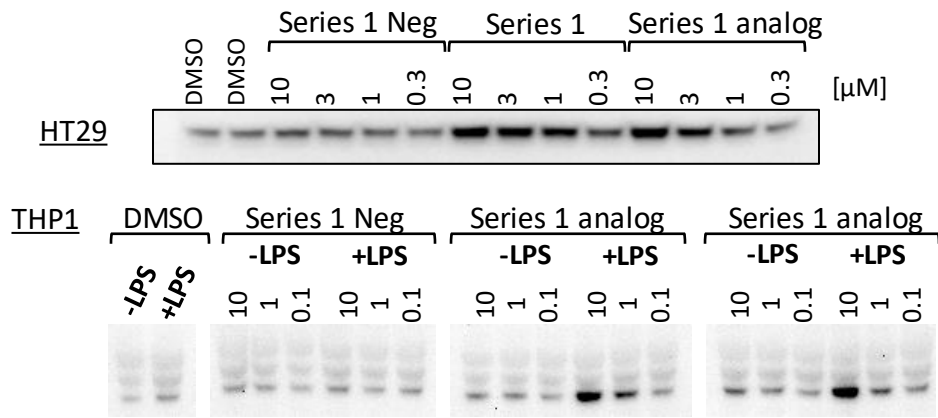


Surface Plasmon Resonance

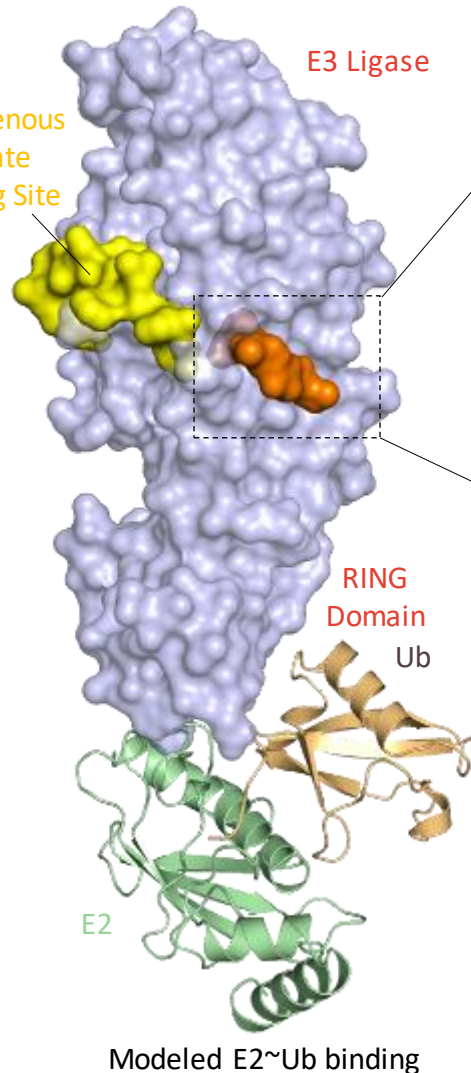


Multiple Linker Vectors Identified from Pellino1 Binders

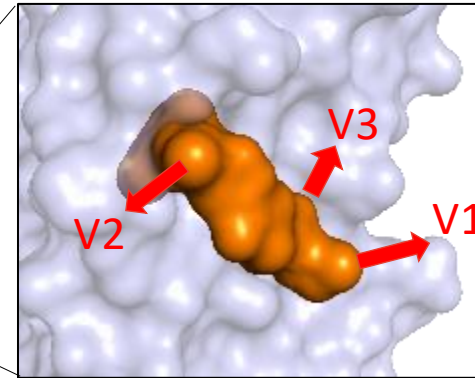
Pellino1 Cellular Levels Increased by Inhibitors



Cells treated with compounds for 24 hours

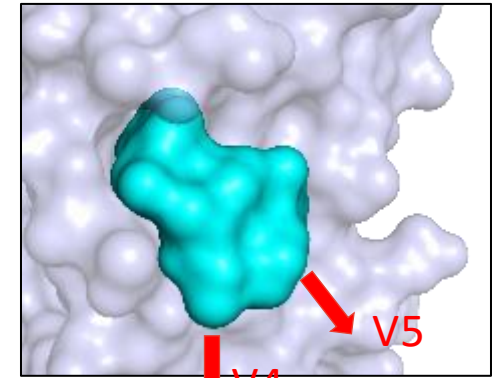


Series 1: Optimized Series Derived from HTS



IC₅₀ = 2 nM

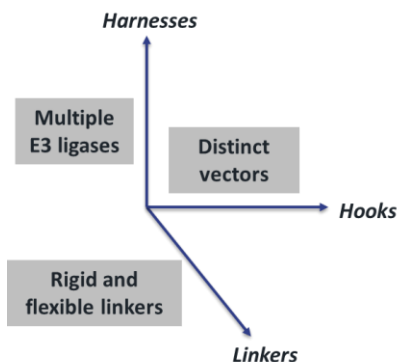
Series 3: Unoptimized DEL Series



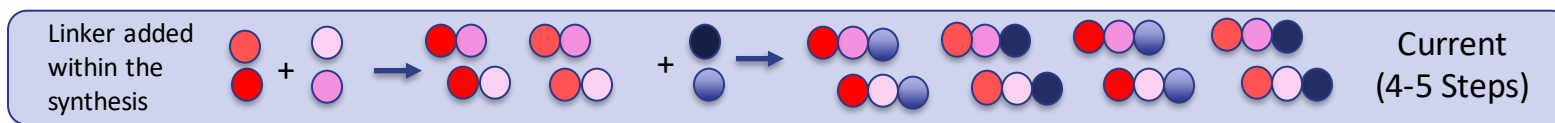
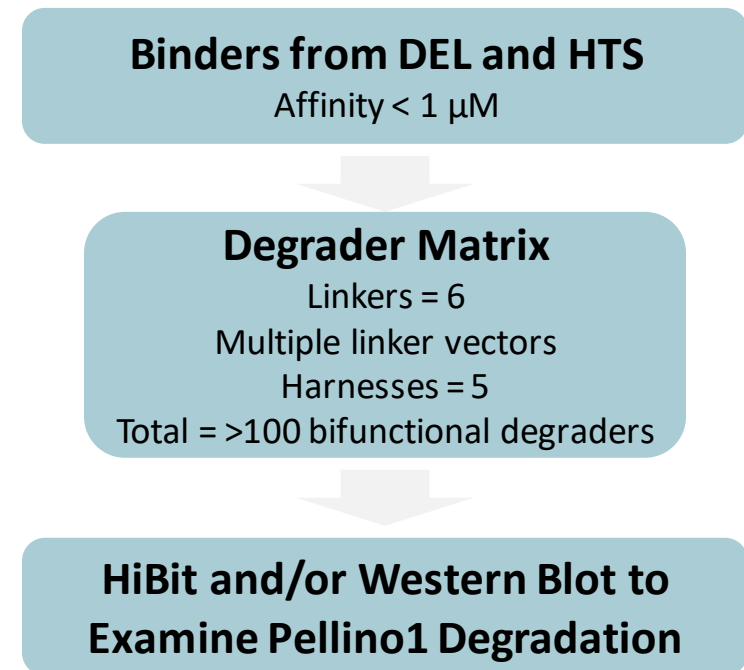
IC₅₀ = 260 nM

- HTS Series was optimized to very potent Pellino1 binder with IC₅₀ = 2 nM
- Unoptimized DEL series identified with IC₅₀ = 260 nM
- Multiple X-ray structure solved of binders in complex with Pellino1
- HTS-derived and DEL-derived series progressed to degrader synthesis to access multiple linker vectors

Matrix Approach to Degradation Hit Identification and Optimization

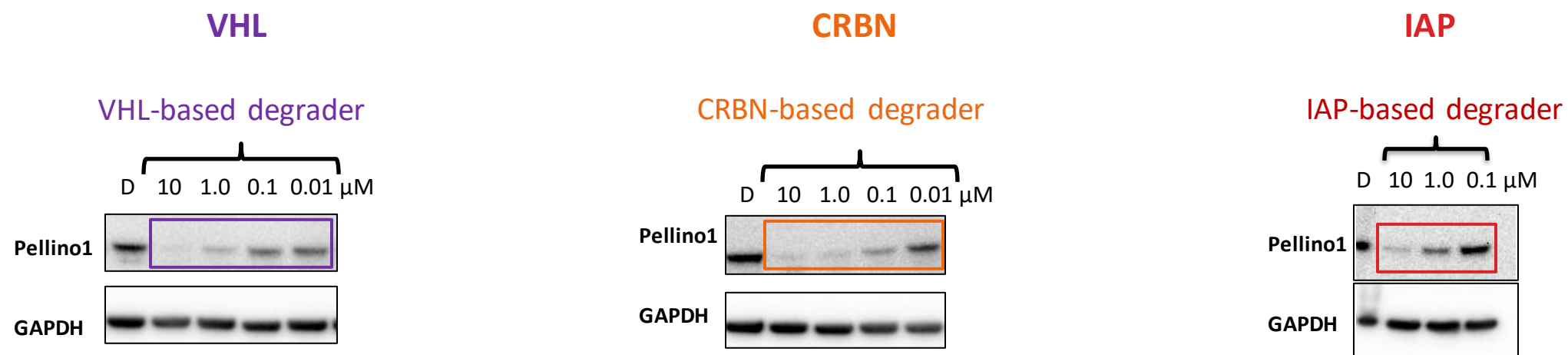


- One compound/well combinatorial libraries
- Up to 5 steps before purification
- Typically, 200-400 degrader compounds made over 4-6 weeks



Diverse combinatorial libraries synthesized

Multiple E3 Ligases Enable Pellino1 Degradation

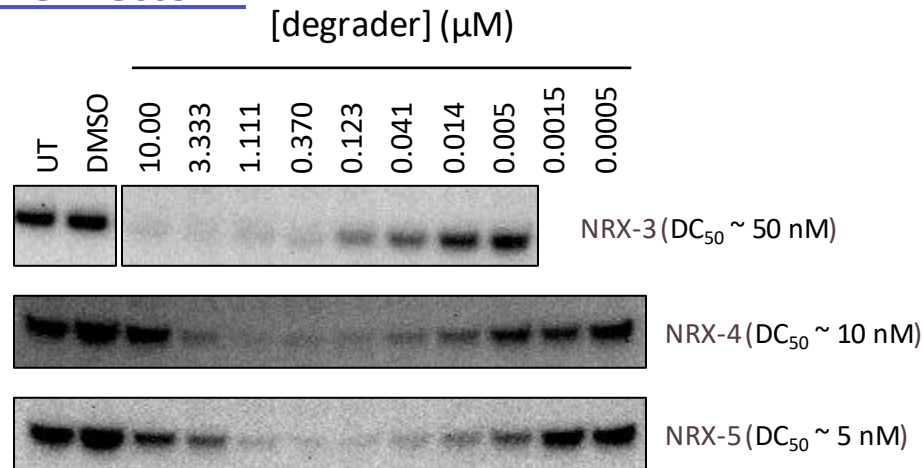


- Multiple ligases identified as being active for Pellino1 degradation
- CRBN-based degraders selected for further exploration

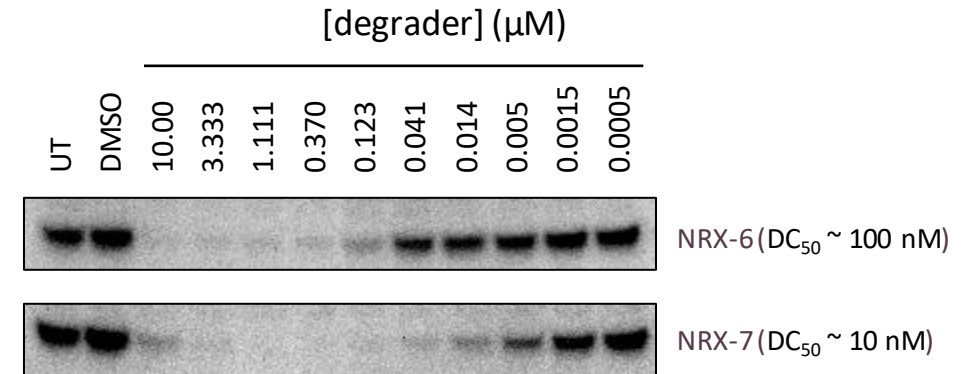
Unstimulated Jurkat cells treated with compound for 24 hours

Pellino1 Degradation Observed with Multiple Linker Vectors

Linker Vector A

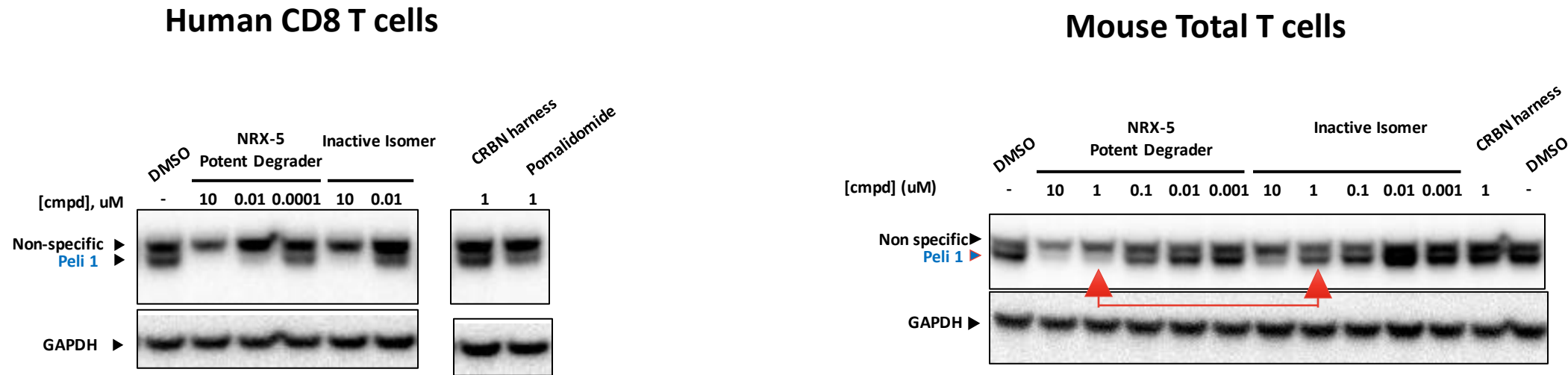


Linker Vector B



- Multiple linker vectors identified which enable potent degradation of Pellino1
- Most potent degraders identified with Linker Vector A
- Other linker vectors resulted in inactive cellular degraders
- Degradation confirmed to be Ubiquitin, NEDD8 and proteasome dependent

Degradation of Pellino1 in Human and Mouse T Cells



- Pellino1 degradation conserved in primary human CD8 T cells and mouse T cells

In Vivo Degradation of Pellino1 in Mice

NRX-5

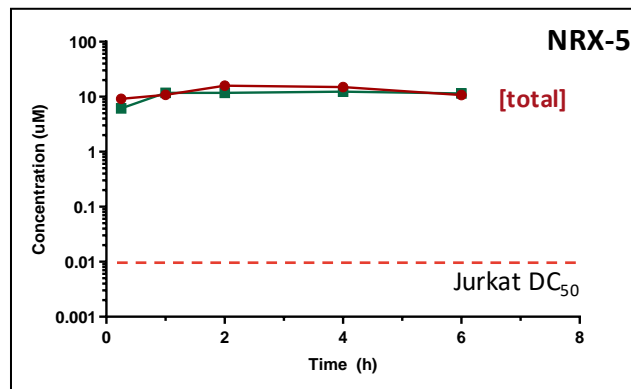
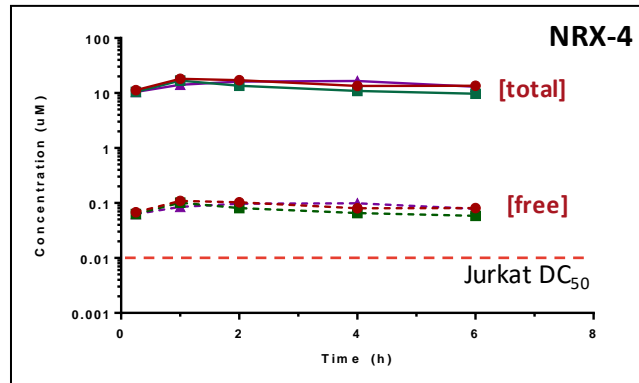
MW = 881

LogD (pH 7.4) = 3.0

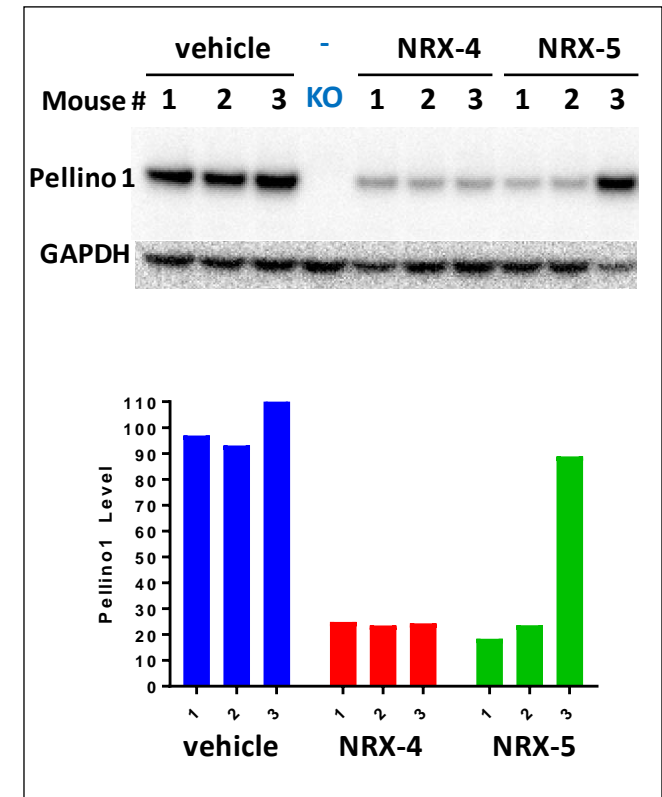
Solubility = 1.4 μ M

Pellino1 DC₅₀ = 10 nM

Single 90 mg/kg IP dose in C57BL/6 Mice



Pellino1 Levels in Mouse Splenocytes (6 hours post single 90 mg/kg IP dose)



Oral Dosing of NRX-8 Demonstrates Pellino1 Degradation

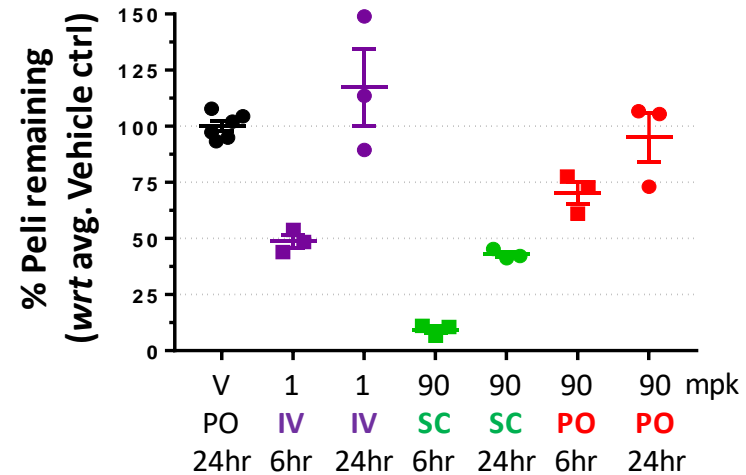
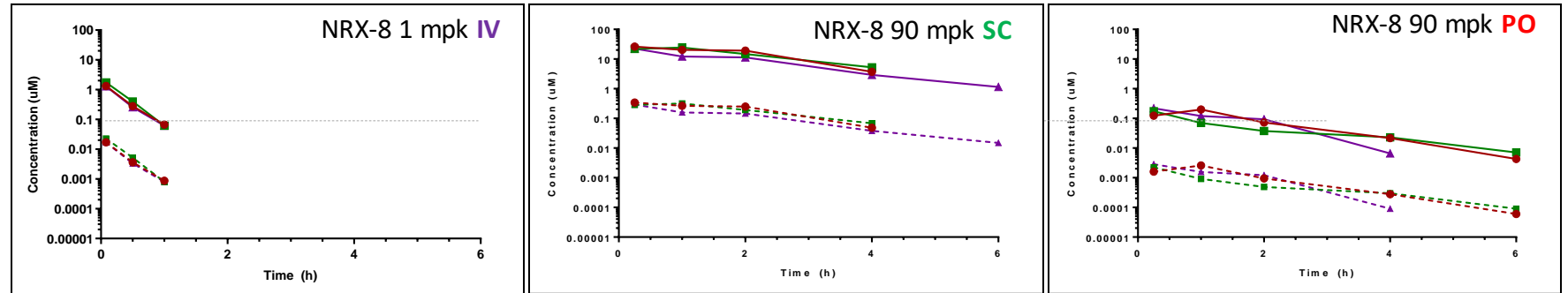
NRX-8

MW = 701

LogD (pH 7.4) = 3.4

Solubility = 7.4 μM

Pellino1 DC_{50} = 2.7 nM



Summary

- Nurix's DELigase platform enables the discovery of potent binders to difficult-to-drug ligase targets with good physicochemical properties
- Matrix approach to identification of active cellular degraders can rapidly yield hit degraders for further optimization
- DELigase platform enabled degradation of Pellino1, an E3 ligase target for immuno-oncology applications
 - Potent cellular degradation of Pellino1 demonstrated with DC50 < 0.1 μ M
 - Cellular degradation preserved across human cell lines, primary human cells and primary mouse cells
 - In vivo degradation of Pellino1 demonstrated in mouse

Thank You

Nurix Therapeutics

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